# A Concise Conversion of (-)-Sclareol into (+)-Coronarin E and (-)-7-*epi*-Coronarin A

Mankil Jung,\* Imju Ko, and Seokjoon Lee

Department of Chemistry, Yonsei University, Seoul 120-749, Korea

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The first synthesis of natural (+)-coronarin E (1) and (-)-7-epi-coronarin A (8) was achieved from (-)-sclareol in four and five steps, respectively.

Labdane terpenes are of special biological interest due to their significant insect antifeedant, antitumor, and antifungal activities, as shown in polygodial,1 coronarin A,2 warburganal,<sup>3</sup> and galanolactone.<sup>4</sup> Coronarin E (1) was isolated from the chloroform extract of the rhizomes of the Brazilian medicinal plant Hedychium coronarium<sup>5</sup> (Zingiberaceae) and Alpinia javanica,6 and from the CH2Cl2 extracts of the aerial parts of Alpinia chinensis.7 Labdienedial (2) and coronarin A are biogenetically related to 1. The furanolabdane coronarin E (1) could be derived by dehydration of 2, while allylic oxidation of 1 could provide coronarin A. There was only one case in which both 1 and **2** had been isolated from the same plant.<sup>6</sup> Consequently, it is likely that 1 is not an artifact, and its presence could be useful as a taxonomic marker.8 Recently, the cytotoxic agent (2) was prepared by Jung et al.<sup>9</sup> No information is available on the synthesis of 1 and coronarin A. Coronarin A, isolated from the rhizomes of *H. coronarium*, showed significant cytotoxic activity (IC<sub>50</sub> = 1.65  $\mu$ g/mL) against Chinese hamster V-79 cells.<sup>2</sup> The interesting biological activities, natural scarcity (0.002% yield), and furanolabdane-type structure of **1** prompted us to synthesize **1** and its related compound **8**. In this note, we report the first synthesis of optically active coronarin E (1) and 7-epicoronarin A (8).

(-)-Sclareol (3) is a readily available natural product whose structure and absolute stereochemistry make it a suitable chiral synthon for the semisynthesis of **1** and **8**.9,10 Thus, oxidative degradation of 3 with osmium tetroxide and sodium periodate in aqueous tert-butyl alcohol cleanly afforded the acetoxyaldehyde 49,10 in one step (65%) (Scheme 1). The acetoxy group at C-8 of compound 4 was derived from the original CH<sub>3</sub> at C-16 of 3.9 Compound 4 could serve as a versatile intermediate for the synthesis of a variety of biologically active natural products, including galanal A,4 warburganal,3 and galanolactone.9 Deacetylation of 4 was regiospecifically achieved with collidine to give **5** (68% yield).<sup>10</sup> Coupling of **5** with 3-bromofuran in the presence of *n*-butyllithium in anhydrous ether at -78°C for 2 h provided the two diastereoisomeric furanolabdane hydroxides 6 in a 3:1 ratio, respectively (yield 71%). Without isomer separation, dehydration of 6 with 2,6lutidine in the presence of methanesulfonyl chloride afforded exclusively 1 (70%). However, dehydration with triethylamine in the presence of methanesulfonyl chloride gave exclusively the cis form (7) (68%). Allylic oxidation of 1 with selenium oxide and tert-butyl peroxide in methylene chloride cleanly provided (-)-7-*epi*-coronarin A (**8**) (73% yield). No natural coronarin A was generated in this oxidation. These syntheses make it possible to solve the problem of natural scarcity of coronarin E (**1**) and (-)-7-*epi*-coronarin A (**8**).

## **Experimental Section**

**General Experimental Procedures.** Melting points were determined in open capillary tubes with a MEL-TEMP II and are uncorrected. IR spectra were obtained as KBr pellets or neat on a NaCl plate, using a Nicolet Impact 400 FT-IR spectrophotometer. The NMR spectra were measured on a Bruker AC 250 NMR spectrometer operating at 250 MHz for <sup>1</sup>H NMR and 63 MHz for <sup>13</sup>C NMR. All spectra were recorded in CDCl<sub>3</sub> as solvent, and chemical shifts were reported in parts per million ( $\delta$ ) relative to TMS. GC/MS spectra were obtained on a Shimadzu GC/MS-QP 2000A. Specific rotations were determined in CHCl<sub>3</sub> using a RUDOLPH Research AUTOPOL III. Analytical TLC was performed on Merck precoated Si gel glass plates (Si gel 60, F<sub>254</sub>, 0.25 mm), and flash column chromatography was performed on Merck 230-400 mesh Si gel. Visualization was achieved with UV (254 nm), solution of molybdic acid (1%) and cerium sulfate (1%) in H<sub>2</sub>SO<sub>4</sub>, anisaldehyde (1%), and H<sub>2</sub>SO<sub>4</sub> (1%) solution in HOAc or KMnO<sub>4</sub> (2%) and NaHCO<sub>3</sub> (4%) solution. Sclareol (3) was purchased from Aldrich Chemical Co., (Milwaukee, WI).

**15,16-Epoxy-12-hydroxy-labda-8(17),13(16),14-triene (6).** A 1.6-M solution of n-butyllithium in hexane (1.2 mL, 1.86 mmol) was added dropwise with stirring to 3-bromofuran (219 mg, 1.49 mmol) in dry ether (10 mL) at -78 °C under nitrogen atmosphere. After 10 min, a solution of **5** (288.3 mg, 1.24 mmol) in dry ether (10 mL) was added into the above reaction mixture. After this mixture had been stirred for 2 h at -78 °C, excess  $H_2O$  was added at room temparature with additional stirring for 30 min. The product was extracted with ether (20 mL  $\times$  3), and the organic layer was dried over MgSO<sub>4</sub> and evaporated. Diastereomers (**6a**:  $R_f$  0.5 and **6b**:  $R_f$  0.3) were separated by Si gel flash column chromatography with eluents (hexane—EtOAc, 5:1) in a 1:3 ratio (**6a**: 69 mg, 18% and **6b**: 205 mg, 54%).

**Compound 6a**: colorless oil; [ $\alpha$ ]<sub>D</sub> +18.54°(c 0.47, CHCl<sub>3</sub>); IR (neat)  $\nu_{\rm max}$  3403, 2933, 2369, 1631, 1469, 1127, 880 cm<sup>-1</sup>; <sup>1</sup>H NMR(CDCl<sub>3</sub>, 250 MHz)  $\delta$  7.38, 7.39 (each 1H, s, H-15, H-16), 6.41 (1H, br s, H-14), 4.86 (1H, s, H-17), 4.69 (1H, dd, J= 8.9, 3.1 Hz, H-12), 4.47 (1H, s, H-17), 0.89, 0.81, 0.68 (each 3H, each s, H-18, 19, 20); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 63 MHz)  $\delta$  149.0 (C-8), 143.2 (C-16), 138.4 (C-15), 130.2 (C-13), 108.5 (C-17), 106.4 (C-14), 65.2 (C-12), 55.3 (C-5), 52.3 (C-9), 42.0 (C-3), 39.2 (C-10), 39.0 (C-1), 38.2 (C-7), 33.6 (C-4), 32.7 (C-11), 24.3 (C-6), 21.7 (C-19), 19.3 (C-2), 14.6 (C-20), 14.1 (C-18); GC/MS m/z 302 (M<sup>+</sup>, 3), 284 (4), 206 (15), 191 (33), 177 (13), 150 (13), 137 (50), 110 (25), 97 (100), 69 (45).

<sup>\*</sup> Tel.: (82)-2-361-2648. Fax: (82)-2-361-2648. E-mail: mkjung@alchemy.yonsei.ac.kr.

#### Scheme 1a

<sup>a</sup> Reagents and conditions: (i) OsO<sub>4</sub> (cat.)/NaIO<sub>4</sub> (1.8 eq.), tert-butyl alcohol, THF, 25 °C, 5.5 h (65%); (ii) collidine, reflux, 25 h, (68%); (iii) 3-bromofuran (1.2 eq.), n-BuLi (1.5 eq.), ether, -78 °C, 2 h., (71%); (iv) MsCl (5 eq.), 2,6-lutidine (10 eq.), CH<sub>2</sub>Cl<sub>2</sub>, room temperature, 30 h (70%); (v) SeO<sub>2</sub> (4 eq.), tert-BuOOH (8 eq.), CH<sub>2</sub>Cl<sub>2</sub>, room temperature, 6 h, (73%).

**Compound 6b**: colorless oil;  $[\alpha]_D + 12.73^\circ$  (*c* 1.5, CHCl<sub>3</sub>); IR (neat)  $\nu_{\text{max}}$  3385, 2933, 1649, 1643, 1125, 890 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz) δ 7.40, 7.34 (each 1H, s, H-15, H-16), 6.42 (1H, br s, H-14), 4.88 (1H, s, H-17), 4.71 (1H, s, H-17), 4.69 (1H, d, J = 5.42 Hz, H-12), 0.82, 0.78, 0.69 (each 3H, each s, H-18, 19, 20); GC/MS m/z 302 (M<sup>+</sup>, 3), 284 (5), 206 (20), 191 (33), 177 (13), 150 (20), 137 (50), 110 (25), 97 (100), 69 (45).

(+)-Coronarin E (1). The base 2,6-lutidine (485 mg, 4.5 mmol) was added to a solution of 6 (273 mg, 0.91 mmol) in dry methylene chloride under N<sub>2</sub> atmosphere at 20 °C with stirring for 30 min. To the stirred reaction mixture was added methanesulfonyl chloride (517 mg, 0.14 mL, 4.5 mmol), and then it was stirred at 20 °C for an additional 18 h. An excess methylene chloride was added to the reaction mixture, which was then washed with 10% HCl solution (20 mL  $\times$  3), saturated NaHCO<sub>3</sub>, and brine, then dried over MgSO<sub>4</sub>. The colorless oily product, 1, was obtained by Si gel column chromatography in 70% yield (176 mg) (eluents: hexane-EtOAc, 5:1): **1** ( $R_f$  0.73), colorless oil;  $[\alpha]_D$  +21.3° (c 0.44, CHCl<sub>3</sub>), lit.<sup>5</sup>[ $\alpha$ ]<sub>D</sub> + 22.3° (c 0.44, CHCl<sub>3</sub>); IR (neat)  $\nu$ <sub>max</sub> 2926, 1604, 1472, 1218, 1128, 1056, 760 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz)  $\delta$  7.36 (2H, s, H-15, H-16), 6.55 (1H, s, H-14), 6.20 (1H, d, J = 15.7 Hz, H-12), 5.98 (1H, dd, J = 15.7 Hz, 9.7 Hz, H-11), 4.76 (1H, s, H-17), 4.54 (1H, s, H-17), 0.90, 0.85, 0.84 (each 3H, each s, H-18, 19, 20);  $^{13}$ C NMR (CDCl<sub>3</sub>, 63 MHz)  $\delta$  150.4 (C-8), 143.0 (C-16), 139.7 (C-15), 128.0 (C-12), 124.7 (C-13), 121.8 (C-11), 107.8 (C-17), 107.6 (C-14), 62.9 (C-9), 54.7 (C-5), 42.5 (C-3), 40.9 (C-1), 39.4 (C-10), 36.8 (C-7), 33.6 (C-4), 33.5 (C-18), 23.4 (C-6), 22.0 (C-19), 19.0 (C-2), 15.1 (C-20); GC/MS m/z 284 (M<sup>+</sup>, 53), 269 (5), 241 (3), 199 (5), 173 (5), 160 (13), 147 (100), 131 (20), 105 (15), 81 (35), 77 (20).

cis-Coronarin E 7. ( $R_f$  0.72), colorless oil; IR (neat)  $\nu_{\rm max}$ 2930, 1594, 1465, 1210, 1132, 1056, 750 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz) δ 7.42, 7.39 (2H, s, H-15, C-16), 6.46 (1H, s, H-14),

4.96 (1H, dd, J = 10.11, 2.15 Hz, H-11), 4.91 (1H, d, J = 10.8Hz, H-12), 4.86 (1H, s, H-17), 4.37 (1H, s, H-17), 0.89, 0.78, 0.69 (each 3H, each s, H-18, 19, 20).

(-)-7-epi-Coronarin A (8). To a stirred solution of SeO<sub>2</sub> (202 mg, 182 mmol) in methylene chloride, tert-BuOOH (656 mg, 7.28 mmol) was added dropwise under a N<sub>2</sub> atmosphere at room temperature. After 10 min, a solution of coronarin E(1) (273 mg, 0.91 mmol) in dry methylene chloride was added with vigorous stirring for 5 h. Excess CH2Cl2 was added to the reaction mixture and washed with H2O, then dried over MgSO<sub>4</sub>. A colorless oily product, 8, was obtained by flash chromatography in 73% yield (eluents: hexane-EtOAc, 5:2): **8** ( $R_f$  0.52), colorless oil;  $[\alpha]_D$  -6.9°(c 0.1, CHCl<sub>3</sub>); IR (neat)  $\nu_{\text{max}}$ 3436, 2933, 1468, 1375, 1204, 1026, 873, 768 cm<sup>-1</sup>; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 250 MHz)  $\delta$  7.35 (2H, s, H-15, H-16), 6.54 (1H, s, H-14), 6.22 (1H, d, J = 15.6 Hz, H-12), 5.97 (1H, dd, J = 15.7 Hz, 9.8 Hz, H-11), 5.00 (1H, s, H-17) 4.71 (1H, s, H-17) 4.42 (1H, d, J = 2.8 Hz, H-7), 2.89 (1H, d, J = 9.6 Hz, H-9), 0.91, 0.84, 0.82 (each 3H, each s, H-18, 19, 20);  $^{13}$ C NMR (CDCl<sub>3</sub>, 63 MHz)  $\delta$ 151.3 (C-8), 143.3 (C-16), 139.8 (C-15), 127.2 (C-12), 124.4 (C-13), 122.4 (C-11), 111.1 (C-14), 107.6 (C-17), 73.4 (C-7), 56.0 (C-9), 47.0 (C-5), 42.2 (C-3), 40.4 (C-1), 39.4 (C-10), 33.3 (C-4), 33.1 (C-18), 30.0 (C-6), 21.8 (C-19), 19.1 (C-2), 14.1 (C-20); GC/ MS m/z 300 (M<sup>+</sup>, 95), 282 (33), 209 (18), 189 (100), 161 (50), 149 (33), 121 (75), 105 (70), 94 (100).

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